

THE EFFECT OF OLIVE OIL ON THE PLASMA TRANSAMINASE ACTIVITIES AND BLOOD HEMATOLOGICAL VALUES OF RATS ACUTELY EXPOSED TO CADMIUM

Branka I. Ognjanović^a, Slađan Z. Pavlović^b, Radoslav V. Žikić^a,
Andraš Š. Štajn^a, Snežana D. Maletić^a, Zorica S. Saičić^b and
Vojislav M. Petrović^b

^aFaculty of Sciences, P.O. Box 60, 34000 Kragujevac, Yugoslavia

^bInstitute for Biological Research "Siniša Stanković", Department of Physiology, 29. Novembra 142, 11060 Belgrade, Serbia, Yugoslavia

(Received April 26, 2000)

ABSTRACT. Male *Wistar albino* 3 months old rats were acutely treated with cadmium (Cd, 0.4 mg/kg b.m., i.p., 24^h before the sacrificing), with olive oil (0.4 mL/kg b.m., i.m., 48^h before the sacrificing) and Cd+olive oil (in above mentioned amounts). The activities of alanin aminotransaminase (ALT) and aspartat aminotransaminase (AST) in the plasma, as well as hematological values: red blood cells (RBC) count, hematocrite (Ht) value, hemoglobin (Hb) and glucose concentrations were determined in the control and experimental groups of animals.

The activities of ALT and AST in the plasma were significantly increased ($p < 0.005$) only in animals treated with cadmium, while in other experimental groups no differences in comparison to controls were found.

Our results show a significant alteration of hematological values. The RBC count ($p < 0.005$), as well as Ht value and Hb concentration ($p < 0.01$) were significantly decreased in respect to the control animals.

Blood glucose concentration was significantly increased ($p < 0.005$) in rats treated with Cd when compared with control rats. However, pretreatment of rats with olive oil diminished the negative effects of cadmium and shows that olive oil prevents or mitigates anemia caused by cadmium.

INTRODUCTION

Cadmium (Cd) is a very toxic heavy metal, an important pollutant of environment (present in soil, water, air, food and in cigarette smoke) which causes poisoning in different organisms (1, 2, 3). Cadmium induces different toxic effects if it is present as a free or non-protein bound (4). After the intake and resorption, Cd enters the blood where it binds to the erythrocyte membranes and proteins of low molecular mass forming metallothioneins (5). Cadmium then transported into the most of tissues and organs in which it also induces the forming of metallothioneins (6, 7, 8, 9). From totally accumulated cadmium in the organism, about 75% is deposited in the liver and kidneys (10, 11, 12). However, Cd is accumulated in most of other tissues and organs, such as pancreas, salivary glands, testes, heart, brain or brown adipose tissue (13, 14, 15, 16, 17, 18).

Binding of cadmium to red blood cells (RBCs) causes their destruction and increased hemolysis, haematological values alterates (decrease of hematocrite value, hemoglobin concentration and total red blood cells count), absorption of intestinal iron is decreased and anemia appears (19, 20, 21). Above mentioned parameters can be taken as the sensitive indicators of cadmium toxicity.

Investigations on different organisms have shown that Cd causes significant metabolic and histological alterations, disturbs biological systems and decreases the body mass growth and mass growth of certain organs (15, 10, 16, 18). Cadmium has negative effects on energy metabolism (20, 22), membrane transport (23, 24) and protein synthesis (25, 10).

Fariss (26) has shown that the scavengers of free radicals and antioxidants may be used in the protection against Cd toxicity. Some antioxidants, such as vitamin E, ascorbic acid, glutathione, selenium and coenzyme Q₁₀ exerts the protective effects against oxidative damages in different tissues of animals treated with cadmium (27, 28, 29, 30, 31).

Mediterranean food is rich in vegetables, cereals, fruits and oil (mostly olive oil), so that the problem of coronary and cardiovascular diseases and arteriosclerosis are diminished in the population of Mediterranean countries. On the basis of many literature data it is established that population of Mediterranean countries which use olive oil in the nutrition in comparison to the countries where the sunflower oil is used, suffer from sicknesses caused by reactive oxygen species (32). Olive oil is composed of polyphenol antioxidants and squalen which decreased production of free radicals and prevent some diseases, especially cardiovascular (33). High content of mono unsaturated fatty acids present in olive oil may also protect organism from the appearance of coronary diseases (34, 35). Olive oil contains lipid antioxidants (vitamin E and β -carotene) which inhibit the oxidation of low density lipoproteins and prevent chain reactions of lipid peroxidation (36).

In this work, possible protective effect of olive oil pretreatment on the plasma transaminase activities and some hematological values of rats acutely treated with cadmium were investigated.

MATERIALS AND METHODS

In our experiments male *Wistar albino* rats, 3 months old (weighing 280 ± 30 g) were used. The animals were kept at $21 \pm 1^\circ\text{C}$ and exposed to 12 h light - 12 h dark cycle. The animals were injected with CdCl₂ (0.4 mg Cd/kg b.m., i.p., 24^h before the sacrificing), with olive oil (0.4 mL/kg b.m., i.m., 48^h before the sacrificing) and with Cd + olive oil (0.4 mg Cd/kg b.m., i.p., 24^h before the sacrificing + 0.4 mL olive oil/kg b.m., i.m., 48^h). Control rats were drunk *ad libitum* by tap water. The exposed rats were housed in individual cages and given a standard diet and water *ad libitum*. Each experimental group consisted of six animals.

The animals were sacrificed by decapitation between 8 and 10 A.M. and fresh blood was immediately collected into heparinized test tubes. RBC count and Ht value were determined by standard hematological techniques (37). The Hb concentration was determined by the cyanmethaemoglobin method (38). The blood glucose concentration was assayed by the ortho-toluidine colorimetric method (39). The activities of ALT and AST in the plasma were determined by spectrophotometric method as suggested by Wooton et al., (40).

Statistical analysis of the results was based on the Student's paired t-test, considering the significance at a level of $p < 0.05$ (41).

RESULTS AND DISCUSSION

Results depicted in Table 1 show a significant increase of the activities of ALT and AST in the plasma ($p < 0.005$) of cadmium exposed rats in comparison with control animals. In other Experimental groups (olive oil and Cd + olive oil), the plasma transaminase activities were similar to those of controls.

The results obtained in this work are in accordance with results of our previous investigations and point out the damage of the liver and disturbed carbohydrate and protein metabolism (42, 43, 44, 28). Similar results were obtained in our investigations on rats after chronic treatment with cadmium (21, 45). Some authors were also shown that cadmium increased the activities of ALT and AST in serum of rabbits (46) and in the plasma of rats (43, 47, 21, 45, 48). These enzymes have an important role in the processes of

aminoacid and protein metabolism. It is known that ALT and AST are widely spread in some tissues and in normal conditions they show very low activity in serum (plasma). However, stress and influence of different pollutants particularly in liver and heart caused the liberation of transaminases into circulation and increased their activities (49). Our results also show that olive oil pretreatment diminished the harmful effects of cadmium on the activities of transaminases in the plasma.

Table 1: The activities of alanin aminotransaminase (ALT) and aspartat aminotransaminase (AST) in the plasma of rats acutely treated with cadmium (Cd), with olive oil and concomitantly treated with Cd and olive oil in respect to the control animals (C).

PLASMA	ALT (U/mL)	AST (U/mL)
C	14.84 ± 0.34	66.31 ± 1.86
Cd	19.91 ± 0.27 ****	94.93 ± 1.78 ****
olive oil	14.20 ± 0.94	68.34 ± 3.88
Cd+olive oil	15.02 ± 0.90	69.83 ± 2.33

Means ± SEM from 6 animals in each group.
Significantly different from controls: ****p<0.005.

RBC count, Ht value, as well as Hb and glucose concentrations were presented in Table 2. Our results show that cadmium treatment induces a significant decrease of RBC count (p<0.005), as well as Ht value and Hb concentration (p<0.01) in respect to the control rats.

Previous investigations show that cadmium induces oxidative damage in erythrocytes, causing destruction of cell membrane and increase lipid peroxidation, as well as the alteration of the antioxidant defence system and energy metabolism and the appearance of anemia (50, 20, 51, 52).

Table 2: Hematological values: red blood cells (RBC) count, hematocrite (Ht) value, hemoglobin (Hb) and glucose concentration in the blood of rats acutely treated with cadmium (Cd), olive oil and concomitantly treated with cadmium and olive oil (Cd+olive oil) in comparison to the control animals (C).

	RBC (10 ¹² /L)	Ht (L/L)	Hb (mmol/L)	Glucose (mmol/L)
C	7.91 ± 0.21	0.45 ± 0.06	8.24 ± 0.11	4.91 ± 0.13
Cd	5.11 ± 0.11 ****	0.41 ± 0.03 ***	7.56 ± 0.10 ***	6.41 ± 0.17 ****
olive oil	7.74 ± 0.15	0.46 ± 0.01	8.42 ± 0.06	5.21 ± 0.15
Cd+olive oil	7.02 ± 0.12 *	0.44 ± 0.01	8.15 ± 0.05	5.17 ± 0.26

Means ± SEM from 6 animals in each group.
Significantly different from controls: * p<0.05, ***p<0.01 and ****p<0.005.

Results obtained in this experiments have confirmed our previous investigations (20, 21) and the results of other authors (19, 50, 51). It is well known, that the presence of cadmium in organism decreases the level of iron in blood (20) and cause the decrease of Hb concentration. The decrease of Ht value in hemolyzed plasma of rats exposed to cadmium indicates the increased destruction of erythrocytes. These results are in accordance with the results of other investigations (19, 50, 20, 51, 48, 52). Cadmium caused the damage of the erythrocyte membrane resulting in hemolysis in the same way like other metals such as Pb,

Cu and Zn (52). Cadmium induced anemia is characterized by pronounced reticulocytosis and hypochromia (20).

In rats exposed to olive oil hematological values were not significantly changed in comparison to the control values. In rats concomitantly treated with cadmium and olive oil the hematological values such as RBC, Ht and Hb were significantly increased in comparison to the values obtained in rats which received cadmium only. Our results suggest a very important role of pretreatment with olive oil before intoxication with cadmium. Olive oil decreased the toxic effects of cadmium on the hematological values and has the protective role in anemia.

The concentration of glucose was increased ($p < 0.005$) in the blood of rats after acute exposure to cadmium, while in rats treated with olive oil and Cd+olive oil the glucose concentration was normalized and was similar to the control values (Table 2).

In animals treated with olive oil before administration of cadmium (pretreated animals) the concentration of glucose was similar to the control values, but it was significantly decreased in comparison to animals which received cadmium only. Pretreatment with olive oil prevents the increase of glucose concentration caused by cadmium.

CONCLUSIONS

Our results show that cadmium causes alterations of carbohydrate and protein metabolism demonstrated as hyperglycemia and increased activities of ALT and AST in the plasma.

Cadmium induced oxidative stress and has an important role in the pathogenesis of anemia and influences oxidative injuries in erythrocytes, their destruction and hemolysis resulting in decreases total red blood cell count, hematocrite value and hemoglobin concentration. These hematological parameters can be taken as sensitive indicators of cadmium toxicity.

It may be concluded, that olive oil exhibit some protective role in the prevention of toxic effects of cadmium on the plasma transaminase activities and some hematological values of rats acutely treated with cadmium.

Acknowledgements: This research was supported by the Ministry of Science and Technology of Serbia, Yugoslavia, Grant Nos. 03E18 and 03E23.

References

- [1] Shukla, G.S., Singhal, R.L., *The present status of biological effects of toxic metals in the environment: lead, cadmium and manganese*. Can. J. Physiol. Pharmacol., 62 (1984), 1015–1031.
- [2] Cifone M.G., Alesse E., Procopio A., Paolini R., Morrone S., Eugenio R.D., Santoni G. and Santoni A., *Effects of cadmium on lymphocyte activation*. Biochem. Biophys. Acta., 1011 (1989), 25–32.
- [3] Stohs, S.J., Bagchi, D., *Oxidative mechanisms in the toxicity of metal ions*. Free Rad. Biol. Med., 18 (1995), 321–336.
- [4] Webb, M., Ed. – *The Chemistry, Biochemistry and Biology of Cadmium*. Amstredam: Elsevier–North Holland Biomedical Press, (1979), pp. 41–422.
- [5] Thornalley, P.J., Vašak M., *Possible role for metallothionein in protection against radiation-induced oxidative stress. Kinetics mechanism of its reaction with superoxide and hydroxyl radicals*. Biochim. Biophys. Acta, 827 (1985), 36–44.
- [6] Ochi, T., Otsuka, F., Takahashi, K., Oshawa, M. *Glutathione and metallothioneins as cellular defense against cadmium toxicity in culture chinese hamster cells*. Chem. Biol. Interactions., 65 (1988), 1–14.
- [7] Viljoen, J.A., Tappel, L.A., *Interactions of selenium and cadmium with metallothionein-like and other cytosolic proteins of rat kidney and liver*. J. Inorg. Biochem., 34 (1988), 277–290.

- [8] Wormser, U., Ben Zakine, S., *Cadmium-induced metallothionein synthesis in the rat liver slice system*. Toxic. in Vitro, 4 (1990), 791–794.
- [9] Nordberg, M., Jin, T., Nordberg, G.F., *Cadmium metallothionein and renal tubular toxicity*. IARC Sci Publ., 118 (1992), 293–297.
- [10] Ognjanović, B., Žikić, R.V., Štajn, A., Saičić, Z.S., Kostić, M.M., Petrović, V.M., *The effects of selenium on the antioxidant defense system in the liver of rats exposed to cadmium*. Physiol. Res., 44 (1995), 293–300.
- [11] Štajn, A., Žikić, R.V., Ognjanović, B., Saičić, Z.S., Pavlović, S.Z., Kostić, M.M., Petrović, V.M., *Effect of cadmium and selenium on the antioxidant defense system in rat kidneys*. Comp. Biochem. Physiol., 117C (1997), 167–172.
- [12] Saygi, S., Deniz, G., Kutsal, O., Vural, N., *Chronic effects of cadmium on kidney, liver, testes and fertility of male rats*. Biol. Trace Elem. Res., 31 (1998), 209–214.
- [13] Kostić, M.M., Ognjanović, B., Žikić, R.V., Štajn, A., Rosić G.L., *Effects of cadmium on antioxidant enzymes, glutathione and lipid peroxidation in brown adipose tissue*. Jugoslav. Physiol. Pharmacol. Acta, 29 (1993a), 137–145.
- [14] Friedman P.A. and Gasek F.A., *Cadmium uptake in the environment*. 2nd Ed. Cleveland, OH: CRC Press., (1994), pp 137–169.
- [15] Štajn, A., Ognjanović, B., Žikić, R.V., Kostić, M.M., *The effect of cadmium on growth of body mass and somatic indexes in young and adult rats*. Coll. Sci. Pap. Fac. Sci. Krag., 13 (1992), 57–63.
- [16] Štajn, A., Žikić, R.V., Ognjanović, B., Plavšić, K., Kostić, M.M., *The effects of selenium on the growth of body mass and somatic indexes in cadmium treated rats*. Conference of selenium, Scientific meetings of the Serbian Academy of Sciences and Arts, Vol LXXVIII, Department of Natural and Mathematical Sciences, 6 (1995), 149–157.
- [17] Swiergosz, R., Zakożenska, M., Sawicka-Kapusta, K., Bacia, K., Jankowska, I., *Accumulation of cadmium and its effect on bank whole tissues after chronic exposure*. Ecotoxicol. Environ. Saf., 41 (1998), 130–136.
- [18] Žikić, R.V., Štajn, A.Š., Ognjanović, B.I., Saičić, Z.S., Kostić, M.M., Pavlović, S.Z., Petrović, V.M., *The effect of cadmium and selenium on the antioxidant enzyme activities in rat heart*. J. Environm. Pathol. Toxicol. Oncol., 17 (1998), 259–264.
- [19] Prigge, E.P., Baumert, H.P., Muhle, H., *Effects of dietary and inhalative cadmium on hemoglobin and hematocrite in rats*. Bull. Environ. Contam. Toxicol., 17 (1977), 585–590.
- [20] Kostić, M.M., Ognjanović, B., Dimitrijević, S., Žikić, R.V., Štajn, A., Rosić, G.L., Ivković, R.V., *Cadmium-induced changes of antioxidant and metabolic status in red blood cells of rats: in vivo effects*. Eur. J. Haematol., 51 (1993b), 86–92.
- [21] Ognjanović, B., *Uticaj kadmijuma i selen na sistem antioksidativne zaštite*. Magistarski rad, Biološki fakultet, Univerzitet u Beogradu. (1993), str. 1–121.
- [22] Petronijević, M.R., Maletić, S.D., Žikić, R.V., Kostić, M.M., *Glycolysis and oxidative pentose phosphate pathway in red blood cells of rats chronically intoxicated with cadmium*. Coll. Sci. Pap. Fac. Sci. Krag., 17 (1995), 191–201.
- [23] Grabowska M. and Guminska M., *Effect of lead and cadmium ions on the ATP-ase activity of the human erythrocyte membrane*. Folia. Med. Cracov, 28 (1987), 123–130.
- [24] Verboost, P.M., Flik, G., Pang, K.T.P., Lock, A.C.R., Bonga, E.W.S., *Cadmium inhibition of the erythrocyte Ca^{2+} pump*. J. Biol. Chem., 264 (1989), 5613–5615.
- [25] Dudley R.E., Svoboda D.J., Kllassi C.D., *Time course of cadmium-induced ultrastructural changes in rat liver*. Toxicol. Appl. Pharmacol., 76 (1984), 150–160.
- [26] Fariss M.W., *Cadmium toxicity: Unique cytoprotective properties of alpha tocopheryl succinate in hepatocytes*. Toxicology, 69 (1991), 63–77.

- [27] Shukla, G.S., Chandra, S.V., *Cadmium toxicity and bioantioxidants: status of vitamin E and ascorbic acid of selected organs in rat*. J. Appl. Toxicol. 9 (1989), 119–122.
- [28] Zamora, R., Hidalgo, F.J., Tappel, A.L., *Comparative antioxidant effectiveness of dietary beta-carotene, vitamin E, selenium and coenzyme Q¹⁰ in rat erythrocytes and plasma*. J. Nutr., 121 (1991), 50–56.
- [29] Hudecova, A., Ginter, E., *The influence of ascorbic acid on lipid peroxidation in guinea pigs intoxicated with cadmium*. Food Chem. Toxicol., 30 (1992), 1011–1013.
- [30] Beyer R.E., *The role of ascorbate in antioxidant protection of biomolecules: Interaction with vitamin E and coenzyme Q*. J. Bioenerg. Biomemb., 26 (1994), 349–358.
- [31] Niki, E., Noguchi, N., Tsuchihashi, H., Gotoh, N., *Interaction among vitamin C, vitamin E, and beta-carotene*. Am. J. Clin. Nutr., 62 (1995), 1322S–1326S.
- [32] Reuter, W., Vorberg, B., Krumpolt, K., Sauer, I., *Effect of olive oil and fish oil on parameters of lipids and antioxidants in hyperlipoproteinemia*. Z. Ernährungswiss, 34 (1995), 151–159.
- [33] Newmark, H.L., *Squalene, olive oil, and cancer risk: a review and hypothesis*. Cancer Epidemiol. Biomarkers Prev., 6 (1997), 1101–1103.
- [34] Mancini, M., Parfitt, V.J., Rubba, P., *Antioxidants in the Mediterranean diet*. Can. J. Cardiol., 11 (1995), 105G–109G.
- [35] Manna, C., Galletti, P., Cucciolla, V., Moltedo, O., Leone, A., Zappia, V., *The protective effect of the olive oil polyphenol (3,4 - dihydroxyphenyl) - ethanol counteracts reactive oxygen metabolite - induced cytotoxicity in Caco-2 cells*. J. Nutr., 127 (1997), 286–292.
- [36] Gerber M., *Olive oil, monounsaturated fatty acids and cancer*. Cancer Lett., 114 (1997), 91–92.
- [37] Chanarin I., *Laboratory haematology. An Account of Laboratory Techniques*. Hong Kong, Churchill Livingstone, (1989).
- [38] Drabkin D. and Austin H., *Spectrophotometric studies preparations from washed blood cells*. J. Biol. Chem., 112 (1935), 51–55.
- [39] Hultmann, E., *Rapid specific method for determination of aldoses in body fluids*. Nature, 183 (1959), 108–109.
- [40] Wooton, E.J., King, E.Y., Yolson, E., Wooton, I.D.P., *In: Micro Analysis in Medical Biochemistry*. 4th ed., London, Churchill, (1964), pp. 1–254.
- [41] Hoel, P.G., *Introduction to mathematical statistics*. John Wiley, New York, (1966), pp. 402–403.
- [42] Chapatwala K.D., Rajanna B. and Desai D., *Cadmium induced changes in gluconeogenic enzymes in rat kidney and liver*. Drug Chem. Toxicol., 3 (1980), 407.
- [43] Rajanna, B., Hobson, M., Reese, J., Sample, E., Chapatwala, K.D., *Chronic hepatic and renal toxicity by cadmium in rats*. Drug Chem. Toxicol., 7 (1984), 229–241.
- [44] Larsson, A., Haux, C., *Altered carbohydrate metabolism in fish exposed to sublethal levels of cadmium*. J. Environm. Biol., 3 (1982), 71–81.
- [45] Štajn, A., Ognjanović, B., Žikić, R.V., Kostić, M.M., *The effect of selenium on the some metabolic processes in cadmium-treated rats*. II Symposium on Chemistry and the Environment, V. Banja, (1993), 283–284.
- [46] Piscator, M., Axelsson, B., *Serum proteins and kidney function after exposure to cadmium*. –Arch. Environ. Health, 21 (1970), 604–608.
- [47] Manca, D., Ricard, A.C., Trottier, B., Chevalier, G., *Studies on lipid peroxidation in rat tissues following administration of low and moderate dose of cadmium chloride*. Toxicology., 67 (1991), 303–323.

- [48] Žikić, R.V., Štajn, A.Š., Ognjanović, B.I., Pavlović, S.Z., Saičić, Z.S., *Activities of superoxide dismutase and catalase in erythrocytes and transaminases in the plasma of carps (Cyprinus carpio L.) exposed to cadmium.* *Physiol. Res.*, 46 (1997), 391–396.
- [49] Henry, R.J., Cannon, D.C., Winkelman, J.W., *Clinical Principles and Technicks.* New York, Harper and Row, (1974), pp. 884–889.
- [50] Kunimoto, M., Miysaka, K., Miura, T., *Changes in membrane properties of rat red blood cells induced by cadmium accumulating in the membrane fraction.* *J. Biochem. Tokyo.*, 99 (1986), 397–406.
- [51] Sarkar, S., Yadav, P., Bhatnagar, D., *Cadmium-induced lipid peroxidation and the antioxidant system in rat erythrocytes: the role of antioxidants.* *J. Trace. Elem. Med. Biol.*, 11 (1997), 8–13.
- [52] Hamada T., Tanimoto A., Arima N., Ide Y., Sasaguri T., Shimajiri S., Murata Y., Wang, K.Y., Sasaguri, Y., *Pathological Study of Splenomegaly Associated with Cadmium-Induced Anemia in Rats.* *Sangyo. Ika. Daigaku. Zasshi.*, 20 (1998), 11–19.